

Claims

What is claimed is:

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- 1. A method of isolating cells comprising,
- (a) obtaining a tissue sample from a subject,
- (b) successively exposing the tissue to a first solution with decreasing amounts of CaCl₂ comprising NaCl, HEPES, MgCl₂, KCl, and sugar at a pH of approximately 7.4,

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- (c) disassociating the tissue with an enzyme solution,
- (d) repeatedly resuspending the disassociated tissue into a second solution with increasing amounts of CaCl₂ comprising Earle's modified salt, L-glutamine, sodium bicarbonate, sodium pentothenate, creatine, taurine, ascorbic acid, HEPES, fetal bovine serum, an antibiotic, and a fatty acid, at a pH of approximately 7.4 to obtain isolated cells.
- 2. The method of claim 1, further comprising the step of resuspending the isolated cells approximately every 24 hours in a solution comprising Earle's modified salt, L-glutamine, sodium bicarbonate, sodium pentothenate, creatine, taurine, ascorbic acid, HEPES, fetal bovine serum, an antibiotic, a fatty acid acid, and CaCl₂ at a pH of approximately 7.4.
- 3. The method of claim 1, further comprising the step of incubating the isolated cells in a mixture of carbon dioxide and air.

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- 4. The method of claim 3, wherein the isolated cells are incubated at approximately 37°C.
- 5. The method of claim 1 wherein, the first solution is exposed to the 30 tissue at approximately 37°C and at approximately 4 ml/min for 3 minutes.
 - 6. The method of claim 1 wherein the concentration of CaCl₂ in the first solution decreases.
- The method of claim 1 wherein the first solution comprises approximately 140 mM NaCl, approximately 10 mM HEPES, approximately 1 mM MgCl₂, approximately 5.4 mM KCl, and approximately 10 mM D-glucose.

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- 8. The method of claim 1 wherein the enzyme solution comprises a digestive enzyme.
- 9. The method of claim 8, wherein the digestive enzyme is a protease or a collagenase.
 - 10. The method of claim 1 wherein the concentration of CaCl₂ in the second solution increases.
- 11. The method of claim 1 wherein the enzyme solution comprises approximately 140 mM NaCl, approximately 10 mM HEPES, approximately 1 mM MgCl₂, approximately 5.4 mM KCl, and approximately 10 mM D-glucose.
- 12. The method of claim 1 wherein the second solution comprises

 Earle's modified salt, L-glutamine, sodium bicarbonate at approximately 1250mg/l, sodium pentothenate, creatine at approximately 328 mg/500ml, taurine at approximately 312mg/500ml, Ascorbic acid at approximately 8.8 mg, HEPES at approximately 2.383g/500ml, fetal bovine serum at approximately 10% v/v, an antibiotic at approximately 5% v/v, a fatty acid at approximately 1 μM at a pH of approximately 7.4.

13. A method of isolating cells comprising,

- (a) obtaining a tissue sample from a subject,
- (b) successively exposing at approximately 37°C the tissue to a first solution with decreasing amounts of CaCl₂ comprising approximately 140 mM NaCl, approximately 10 mM HEPES, approximately 1 mM MgCl₂, approximately 5.4 mM KCl, and approximately 10 mM sugar at a pH of approximately 7.4,
 - (c) disassociating the tissue with an enzyme solution for approximately 8 minutes comprising approximately 140 mM NaCl, approximately 10 mM HEPES, approximately 1 mM MgCl₂, approximately 5.4 mM KCl, and approximately 10 mM sugar, to form disassociated cells,
 - (d) repeatedly resuspending the disassociated cells into a second solution with increasing amounts of CaCl $_2$ comprising Earle's modified salt, L-glutamine, sodium bicarbonate at approximately 1250mg/l, sodium pentothenate, creatine at approximately 328 mg/500ml, taurine at approximately 312mg/500ml, ascorbic acid at approximately 8.8 mg, HEPES at approximately 2.383g/500ml, fetal bovine serum at approximately 10% v/v, an antibiotic at approximately 5% v/v, and a fatty acid at approximately 1 μ M at a pH of approximately 7.4 to form a solution of isolated cells,

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(e) incubating the isolated cells in a mixture of carbon dioxide and air at approximately 37°C, and

(f) re-suspending the isolated cells approximately every 24 hours in a solution comprising Earle's modified salt, L-glutamine, sodium bicarbonate, sodium pentothenate, creatine, taurine, ascorbic acid, HEPES, fetal bovine serum, an antibiotic, a fatty acid, and CaCl₂ at a pH of approximately 7.4 to obtain isolated cells.

- 14. A method of cultivating isolated cells comprising, resuspending the isolated cells approximately every 24 hours in a solution comprising Earle's modified salt, L-glutamine, sodium bicarbonate, sodium pentothenate, creatine, taurine, ascorbic acid, HEPES, fetal bovine serum, an antibiotic, a fatty acid, and CaCl₂ at a pH of approximately 7.4.
- The method of claim 14 wherein the solution comprises sodium bicarbonate at approximately 1250mg/l, creatine at approximately 328 mg/500ml, taurine at approximately 312 mg/500ml, ascorbic acid at approximately 8.8 mg/500 ml, HEPES at approximately 2.383 g/500ml, fetal bovine serum at approximately 10% v/v, an antibiotic at approximately 5% v/v, and a fatty acid at approximately 1 μM, and approximately 1 mM CaCl₂.

16. A cell culture media for cells comprising Earle's modified salt, L-glutamine, sodium bicarbonate, sodium pentothenate, creatine, taurine, ascorbic acid, HEPES, fetal bovine serum, an antibiotic, a fatty acid, and CaCl₂ at a pH of approximately 7.4.

17. The cell culture media of claim 16 wherein the media comprises sodium bicarbonate at approximately 1250mg/l, creatine at approximately 328 mg/500ml, taurine at approximately 312 mg/500ml, ascorbic acid at approximately 8.8 mg/500 ml, HEPES at approximately 2.383 g/500ml, fetal bovine serum at approximately 10% v/v, an antibiotic at approximately 5% v/v, a fatty acid at approximately 1 μ M, and approximately 1 mM CaCl₂.

- 18. A method of isolating cells comprising,
- (a) obtaining a tissue sample comprising cells from a subject;
- (b) chopping the tissue;
- (c) incubating the tissue in a first solution comprising calcium, salts, magnesium sulfate, pyruvate, glucose, taurine, HEPES, and nitrilotriacetic acid;

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- (d) incubating the tissue in a second solution comprising calcium, salts, magnesium sulfate, pyruvate, glucose, taurine, HEPES, and a digestive enzyme;
- (e) incubating the tissue in a third solution comprising calcium, salts, magnesium sulfate, pyruvate, glucose, taurine, HEPES, and a digestive enzyme; and
 - (f) centrifuging the tissue to obtain isolated cells.
- The method of claim 18, further comprising the step of 19. resuspending the isolated cells in a culture media comprising medium M199, BSA, ascorbic acid, taurine, carnitine, creatinine, insulin, and an antibiotic .
- 20. The method of claim 19, wherein the culture media further comprises a fatty acid or magnesium.
- The method of claim 18, wherein the first solution comprises 21. approximately 1-2 μM CaCl₂, approximately 120mM NaCl, approximately 5.4 mM KCl 15 5.4, approximately 5 mM MgSO₄, approximately 5 mM pyruvate, approximately 20 mM glucose 20, approximately 20 mM taurine, approximately 10 mM HEPES, and approximately 5 mM nitrilotriacetic acid, at a pH of approximately 6.96.
- 20 The method of claim 18, wherein the second solution comprises 22. approximately 1-2 μM CaCl2, approximately 30 μM NaCl, approximately 5.4 mM KCl 5.4, approximately $5\ \text{mM}\ \text{MgSO}_4$, approximately $5\ \text{mM}$ pyruvate, approximately $20\ \text{mM}$ glucose 20, approximately 20 mM taurine, approximately 10 mM HEPES, and 4 U/ml of a digestive enzyme. 25
 - 23. The method of claim 18, wherein the third solution comprises approximately 1-2 μM CaCl2, approximately 30 μM NaCl, approximately 5.4 mM KCl 5.4, approximately 5 mM MgSO₄, approximately 5 mM pyruvate, approximately 20 mMglucose 20, approximately 20 mM taurine, approximately 10 mM HEPES, and 4 U/ml of a digestive enzyme.
 - A method of isolating cells comprising, 24.
 - (a) obtaining a tissue sample comprising cells from a subject;
 - (b) chopping the tissue;
- 35 (c) incubating the tissue in a first solution comprising approximately 1-2 μM CaCl₂, approximately 120mM NaCl, approximately 5.4 mM KCl 5.4, approximately 5 mM MgSO₄, approximately 5 mM pyruvate, approximately 20 mM glucose 20,

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approximately 20 mM taurine, approximately 10 mM HEPES, and approximately 5 mM nitrilotriacetic acid, at a pH of approximately 6.96;

- (d) shaking the tissue at approximately 37°C for approximately 12 minutes;
- (e) bubbling approximately $100\% O_2$ through the solution;
- (f) incubating the tissue in a second solution comprising approximately 1- $2~\mu M~CaCl_2,$ approximately 30 $\mu M~NaCl,$ approximately 5.4 mM KCl 5.4, approximately 5 mM MgSO₄, approximately 5 mM pyruvate, approximately 20 mM $\,$ glucose 20, approximately 20 mM taurine, approximately 10 mM HEPES, and 4 U/ml of a digestive enzyme;
- (g) incubating the solution in a third solution comprising third solution comprises approximately 1-2 μM CaCl2, approximately 30 μM NaCl, approximately 5.4 mM KCl 5.4, approximately 5 mM MgSO₄, approximately 5 mM pyruvate, approximately 20 mM glucose 20, approximately 20 mM taurine, approximately 10 mM HEPES, and 4 U/ml of a digestive enzyme; and
 - (h) centrifuging the tissue to obtain isolated cells.
- A method of isolating and cultivating human myocardial cells 25. comprising,
- (a) obtaining a tissue sample comprising myocardial cells from a human subject;
 - (b) chopping the tissue;
- (c) incubating the tissue in a first solution comprising approximately 1-2 μM calcium, approximately 120mM NaCl, approximately 5.4 mM KCl, approximately 5 mM MgSO₄, approximately 5 mM pyruvate, approximately 20 mM glucose 20, 25 approximately 20 mM taurine, approximately 10 mM HEPES, and approximately 5 mM $\,$ nitrilotriacetic acid, at a pH of approximately 6.96;
 - (d) shaking the tissue at approximately 37°C for approximately 12 minutes;
 - (e) bubbling approximately 100% O2 through the solution;
 - (f) incubating the tissue in a second solution comprising approximately 1- $2~\mu\text{M},$ approximately 30 μM NaCl, approximately 5.4 mM KCl, approximately 5 mM MgSO₄, approximately 5 mM pyruvate, approximately 20 mM glucose 20, approximately 20 mM taurine, approximately 10 mM HEPES, and 4 U/ml of a digestive enzyme;
 - (g) incubating the solution in a third solution comprising third solution comprises approximately 1-2 $\mu M,$ approximately 30 μM NaCl, approximately 5.4 mM KCl 5.4, approximately 5 mM MgSO₄, approximately 5 mM pyruvate, approximately 20

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mM glucose 20, approximately 20 mM taurine, approximately 10 mM HEPES, and 400 U/ml of a digestive enzyme;

- (h) centrifuging the tissue to obtain isolated cells;
- (i) repeatedly resuspending the disassociated cells into a second solution which comprises increasing amounts of CaCl₂, Earle's modified salt, L-glutamine, sodium bicarbonate at approximately 1250mg/l, sodium pentothenate, creatine at approximately 328 mg/500ml, taurine at approximately 312mg/500ml, ascorbic acid at approximately 8.8 mg, HEPES at approximately 2.383g/500ml, fetal bovine serum at approximately 10% v/v, an antibiotic at approximately 5% v/v, and a fatty acid at approximately 1 μM at a pH of approximately 7.4 to form a solution of isolated cells; and
 - (j) incubating the isolated cells in a mixture of carbon dioxide and air at approximately 37°C.
 - 26. A method of isolating and cultivating rodent myocardial cells comprising,
 - (a) removing the heart of a rodent;
 - (b) perfusing the heart with low calcium Tyrode's solution for approximately 3 minutes;
 - (c) perfusing the heart with an enzymatic solution for approximately 8 minutes;
 - (d) perfusing the heart with a low calcium solution for approximately 3 minutes;
 - (e) removing the ventricles;
 - (f) mincing the ventricles to isolate myocardial cells;
 - (g) mixing the cells in a low calcium solution;
 - (h) resuspending the cells in a solution comprising increasing concentrations of calcium; and
 - (i) resuspending the cells in culture media solution..

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